Treatment of brain tumors in children is associated with abnormal MRS ratios in brain tissue remote from the tumor site.

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Treatment of Brain Tumors in Children Is Associated with Abnormal MR Spectroscopic Ratios in Brain Tissue Remote from the Tumor Site

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PURPOSE: Children who have brain tumors are at risk for a variety of treatment-related sequelae, including neuropsychological and cognitive impairment, neurologic deficits, and neuroendocrinologic disturbances. We sought to determine the value of proton MR spectroscopy in assessing brain tissue remote from the tumor site to ascertain the effects of chemotherapy and radiation treatment in these patients.

METHODS: Single-voxel proton MR spectra from 70 patients (111 spectra) and 11 healthy volunteers (11 spectra) were analyzed. NAA/Cr, NAA/Cho, and Cho/Cr ratios based on peak areas were obtained from nonneoplastic regions of the frontal lobe. The relationship between MR spectroscopic ratios and treatment was determined.

RESULTS: NAA-containing ratios were decreased in patients as compared with control subjects. The presence of gadolinium-based contrast material did not cause significant changes in the ratios as compared with precontrast data. When chemotherapy was a component of a child's treatment protocol, we found a significant decline in NAA/Cr ratios. Patients who underwent both chemotherapy and radiation therapy showed a trend toward lower NAA-containing ratios if the chemotherapy was administered before the radiation therapy. Patients receiving whole-brain radiation had a trend toward lower NAA-containing ratios than did those who had only focal tumor treatment.

CONCLUSION: In children with brain tumors, MR spectroscopy of brain tissue remote from the tumor reveals treatment-related biochemical changes.

Children who have brain tumors are at risk for a variety of treatment-related sequelae, including neuropsychological and cognitive impairment, neurologic deficits, and neuroendocrinologic disturbances (1–4). These may result from brain injury associated with the primary tumor itself, from surgical intervention, or from the effects of radiation therapy and chemotherapy on nontumorous brain tissue (5–8). Among the better-known adverse effects are those of radiation therapy, whether used alone or in combination with chemotherapy, with resultant leukoencephalopathy, vasculopathy, or tissue necrosis (8–15). The adverse effects of brain tumor treatment tend to worsen with time, with tumor relapse, and with multiple or aggressive therapies. Additionally, these effects are accentuated in children who are very young at the time of diagnosis (2).

While CT and MR imaging provide excellent anatomic information, there is a need for noninvasive physiological measurements that can serve as more sensitive markers for treatment-related effects on normal tissue. Proton MR spectroscopy provides biochemical information that can potentially affect treatment planning or prompt early intervention to prevent cognitive impairment.

MR spectroscopy uses MR technology to obtain additional information that is unavailable from anatomic MR imaging (16, 17). The signals typically studied with MR spectroscopy include moieties contain-
ing N-acetyl aspartate (NAA), a neuronal marker; choline (Cho), a marker of membrane integrity; creatine (Cr), a bioenergetic metabolite; and lactate, a marker of anaerobic metabolism (16, 17). Although experience to date is limited, other investigations have suggested a role for MR spectroscopy in the noninvasive assessment of tumor type and malignancy, tissue necrosis, ischemia, demyelination, and the effects of radiation therapy (18–26).

To investigate the role of MR spectroscopy as a marker for treatment-related effects, we successfully obtained 111 spectra from uninvolved brain tissue remote from the primary tumor site in 70 children with primary brain tumors, and 11 spectra from similar positions in 11 healthy pediatric volunteers. MR spectroscopic ratios (NAA/Cho, NAA/Cr, and Cho/Cr) were then compared with clinical parameters based on treatment protocols.

Methods

Subjects

MR spectroscopy was added to the clinically indicated MR imaging studies of 81 patients being followed up for primary brain neoplasms, resulting in 164 available spectra. Fifty-three MR spectroscopic examinations and 11 patients were excluded from further study owing to variable voxel placement early in our experience with MR spectroscopy, failure of automated MR spectroscopic shimming and water suppression, poor MR signal-to-noise ratio, or insufficient signal-to-noise and peak resolution for peak area measurements. Therefore, patient data from 111 spectra (70 patients, ages 2 to 22 years; mean age, 10.9 ± 5.0) and control data from 11 spectra (11 healthy volunteers, ages 7 to 15 years; mean age, 10.4 ± 2.9) are included in this report. In addition to these studies, comparison pre- and postcontrast spectra were obtained from the same frontal lobe location in 10 patients to establish the effect of paramagnetic contrast material on MR spectroscopic data. Informed consent as required by our institutional human investigation guidelines was obtained for all patients and volunteers. Children with braces, dental implants, preexisting neurologic or genetic conditions unrelated to tumor, insufficient sedation, or lack of cooperation for MR spectroscopic acquisition were excluded from the study.

MR Spectroscopic Data Acquisition and Processing

Spectra were acquired using the standard head coil on a 1.5-T system equipped with an automated program for single-voxel MR spectroscopic studies (PROBE/SV: GE Medical Systems; Milwaukee, WI). Single MR spectroscopic voxels (5 to 8 mm³) containing a mixture of white and gray matter were located within the right or left frontal lobe remote from the tumor site at the level of the genu of the corpus callosum as determined from the midline sagittal scout image. Voxels were located graphically from an axial plane image (Fig 1). Spectra were acquired using a point-resolved spectroscopic (PRESS) sequence with parameters of 2000/270 (TR/TE), 64 or 128 acquisitions, 2048 data points, and a total examination time of 5 to 10 minutes per voxel. The gradient order was changed from the default order of zyx to yxz to compensate for inhomogeneities associated with voxel proximity to the paranasal sinuses (27). Baseline shimming of the magnet was performed approximately every 6 months to correct for excess automated shimming failures. Localized shimming of the voxel was performed before every examination.

Because paramagnetic contrast material was necessary for the neuroimaging portion of the study, the MR spectroscopic examinations in patients were completed within 25 minutes after administration of the contrast agent. Control subjects did not receive contrast material. Nineteen patients had additional spectra taken from the contralateral frontal lobe using identical MR spectroscopic parameters. Four patients had spectra from only the left frontal lobe owing to abnormalities in the right frontal lobe. To better determine the effect of paramagnetic contrast material on MR spectroscopic data, 10 patients had additional MR spectra taken before contrast administration.

MR spectroscopic data were processed on a workstation using Sage software (GE Medical Systems, Milwaukee, Wis), which included an automated sequence for the processing of PROBE data as described by Webb et al (28). The automated processing is designed to correct receiver-phase and phase errors due to eddy currents. The areas of spectral peaks representing NAA, Cho, and Cr were determined by digitizing the processed spectra. A baseline was estimated and the peaks were traced on a digitizing pad (Jandel Scientific Software, Chicago) interfaced to a computer running Sigma Scan (Jandel). NAA/Cr, NAA/Cho, and Cho/Cr ratios were calculated from these peak areas.

Because some patients were examined on multiple occasions during the course of this study, patients with repeat MR spectroscopic measurements without a change in treatment score were represented by an averaged value in the statistical calcu-
lations (n = 21 patients). Patients whose treatment score changed at the time of follow-up MR spectroscopy (n = 9) were represented more than one time in the statistical analysis that follows. Eight patients are represented by two scores each and one patient by three scores.

Clinical Data and Ratings

Chart review was completed for tumor type, age at diagnosis, tumor location, treatment history, clinical course, and MR imaging findings. Two experienced investigators who were blinded to the MR spectroscopic results assigned each child a consensus treatment score for each MR spectroscopic measurement, with points assigned as follows: presence of a tumor = 1, surgical resection = 1, radiation therapy = 1 per treatment course, and chemotherapy = 1 per treatment course. Control subjects received a score of zero. Because the treatment regimens encountered within this study varied considerably in dose and type of radiation therapy or chemotherapy, a detailed analysis of the effects of individual protocols of radiation therapy or chemotherapy was not attempted. Two items related to treatment protocol were examined for effects on MR spectroscopic ratios: the effect of whole-brain radiation versus focal tumor radiation therapy alone and the effect of order of administration of chemotherapy and radiation therapy. Chemotherapeutic agents represented in our patients' treatment regimens included vincristine, etoposide (VP-16), cyclophosphamide, cisplatin, ifosfamide, 5-fluorouracil, and carmustine. Radiation therapy courses were recorded by portal location as whole-brain and/or focal tumor therapy and by dose administered. Three children had only whole-brain treatment (4000 to 5400 cGy; mean, 4813 ± 727), 15 had only focal tumor portals (4500 to 5400 cGy; mean, 5109 ± 249) including one with an unknown dose, and 30 children had both whole-brain (2600 to 5040 cGy; mean, 3698 ± 589 cGy) and focal tumor (3600 to 2400 cGy; mean, 1544 ± 494 cGy) therapy with a total dose of 4100 to 6720 cGy (mean, 5243 ± 526). Seven patients had radiation therapy at other institutions; thus, complete treatment details were not available.

Statistical Analysis

Statistical analyses were performed using SAS (SAS Institute, Cary, NC) and SPSS (SPSS, Inc, Chicago) statistical programs and included t-tests for independent and paired samples, Levene's test for equality of variances, one-way ANOVA, and Tukey analysis. The relationship was determined between MR spectroscopic ratios (NAA/Cr, NAA/Cho, and Cho/Cr) and the following: 1) left versus right frontal lobe voxel location; 2) presence or absence of paramagnetic contrast administration; 3) treatment score; 4) surgical resection, radiation therapy, or chemotherapy as components of a treatment protocol; 5) whole-brain radiation therapy (n = 3 whole brain and n = 30 whole brain plus focal tumor) versus focal tumor treatment alone; 6) relative order of chemotherapy and radiation therapy; and 7) tumor type. Probability levels of .05 were considered significant. Mean values are reported with standard deviations.

Results

The types of brain tumors found in the children in this study included primitive neuroectodermal tumor (PNET, n = 23), low-grade astrocytoma (n = 31), malignant astrocytoma or glioblastoma multiforme (n = 3), germ cell or germinoma (n = 4), craniopharyngioma (n = 2), ependymoma (n = 5), malignant melanoma (n = 1), and hemangiopericytoma (n = 1).
Children who received chemotherapy before radiation therapy had a trend toward lower NAA/Cr and NAA/Cho ratios than seen in children receiving radiation therapy only or radiation before chemotherapy (Table 3). Additionally, NAA/Cr and NAA/Cho ratios in the chemotherapy-first group were significantly lower \( (P < .05) \) than those of the control subjects.

As expected, ratios from children whose therapy included whole-brain radiation showed a trend toward lower NAA/Cr and NAA/Cho values as compared with children treated with focal tumor radiation therapy (Table 3). Children having whole-brain radiation therapy also had NAA/Cr and NAA/Cho ratios significantly lower than those of control subjects (Table 3). There was a significant linear relationship between whole-brain radiation dose and NAA/Cr \( (r = -.67, P < .05) \) in the 11 patients who did not have chemotherapy (Fig 3A). In patients who had both whole-brain radiation and chemotherapy \( (n = 22) \), there was no such linear trend \( (r = .24) \) (Fig 3B). Treatment via surgical resection had no significant effect on the ratios (Table 3).

Relationship between MR Spectroscopy and Tumor Type

A comparison of MR spectroscopic values by tumor type showed children with PNET to have NAA/Cr values significantly lower than those of control subjects \( (P < .05) \). Only the PNET, low-grade astrocytoma, and ependymoma groups were large enough for comparison (Table 4).

Discussion

Proton MR spectroscopy has proved helpful for studying brain tumor metabolism and for distinguishing tumor recurrence from radiation necrosis (21–24). Our study shows that single-voxel MR spectroscopy can be used successfully as an adjunct to MR imaging in the examination of treatment effects on uninvolved brain in children with primary brain tumors. In this study, MR spectroscopic ratios were obtained from a frontal lobe voxel that contained both white and gray matter. The voxel location was selected to include a substantial component of white matter, owing to the
known adverse effects of radiation and chemotherapy
on white matter. Because we were studying a pediat-
ric population, we expected a disproportionate num-
ber of primary tumors to be located in the posterior
fossa. Our expectation was that treatment effects on
nontumorous brain tissue could best be studied by
using a voxel as anatomically distant from the primary
tumor site as possible. Finally, since young children
were included in the study, the larger size of the
frontal lobe and ease of voxel positioning in younger
children were advantageous. To offset the adverse
effects of magnetic susceptibility artifacts from the
paranasal sinuses on frontal lobe MR spectroscopic
data, the gradient order was altered as described by
Ernst and Chang (27).

The variability in left versus right frontal lobe MR
spectroscopic ratios in this study was 20% (SD/
mean × 100), well within the range of 18% to 38%
variability that has been reported in other clinical MR
spectroscopic studies (23, 37) and less than the frontal
lobe variability recently reported by Jayasundar and
Raghunathan (29). Although multisite automated
single-voxel MR spectroscopic data from healthy
adult volunteers have shown lower variability (10%)
in MR spectroscopic ratios (28), to our knowledge
this has not been achieved in patient studies. Factors
affecting variability may include precision of voxel
placement, magnetic susceptibility artifacts from the
adjacent paranasal sinuses, subject motion, biological
variation, and disease state (for patients). Our study
indicates that owing to the variability of MR spectro-
scopict measures, MR spectroscopic values are most
useful as markers of treatment-related effects across
a group of patients in whom multiple and
longitudinal MR spectroscopic measurements are
available. Single-patient or individual MR spectro-
scopic measurements should be used only with cau-
tion in conjunction with other clinical and imaging
parameters as markers of treatment-associated ef-
scts in a given patient.

MR spectroscopic acquisitions using a long TE
(270 milliseconds) were selected to maximize depic-
tion of NAA, Cr, Cho, and lactate spectral peaks.

### TABLE 2: Mean MR spectroscopic ratios for all control subjects and
treatment scores

<table>
<thead>
<tr>
<th>No.</th>
<th>NAA/Cr</th>
<th>NAA/Cho</th>
<th>Cho/Cr</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control subjects</td>
<td>NAA/Cr</td>
<td>2.77 ± 0.59</td>
<td>2.10 ± 0.43</td>
</tr>
<tr>
<td>Patients</td>
<td>NAA/Cho</td>
<td>2.31 ± 0.47*</td>
<td>1.80 ± 0.37*</td>
</tr>
<tr>
<td>Treatment score</td>
<td>Cho/Cr</td>
<td>2.25 ± 0.25</td>
<td>2.07 ± 0.81</td>
</tr>
<tr>
<td>1</td>
<td>7</td>
<td>2.55</td>
<td>1.88</td>
</tr>
</tbody>
</table>

* Ratios were significantly lower for all patients than for control subjects (t-test, \( P < .05 \)).
† NAA/Cr ratios were significantly lower in these patients than in control subjects (Tukey analysis, \( P < .05 \)).

Brain tumor therapy has been associated with tissue
changes resulting from ischemia, ischemic demyelina-
tion, and neuronal loss. Thus, we hoped that these
would be apparent in the MR spectroscopic ratio
changes.

Most studies of MR spectroscopy in tumor assess-
ment use spectral data obtained from the same pa-
tient’s nontumorous brain, contralateral or adjacent
to the tumor site, for the normal comparison. Our
study shows that there are measurable MR spectro-
scopict differences in nontumorous brain tissue
remote from the tumor site, possibly invalidating the
assumption that spectra obtained from nontumorous
brain parenchyma are “normal.” Instead, it is likely
that MR spectroscopic measurements from untreated
brain that are acquired before tumor therapy treat-
ment are more accurate for use as control or baseline
values.

As expected, MR spectroscopic ratios from nontu-
morous brain tissue grouped by primary tumor type
revealed few tumor-specific findings. The small size
of many of the tumor groups prevented us from fur-
ther analysis of this parameter.

Because we identified treatment-specific decreases
in both NAA/Cho and NAA/Cr (Table 3) in the
patient subgroups, our findings suggest that NAA is
the major metabolite detectable by MR spectroscopy
in nontumorous brain tissue affected by brain tumor
therapy. Although NAA is considered to be a strong
neuronal marker, its presence has been confirmed in

### TABLE 3: Effect of treatment methods on MR spectroscopic ratios

<table>
<thead>
<tr>
<th>Treatment</th>
<th>No. of Patients</th>
<th>NAA/Cr</th>
<th>NAA/Cho</th>
<th>Cho/Cr</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chemotherapy</td>
<td>No</td>
<td>48</td>
<td>2.42 ± 0.51</td>
<td>1.82 ± 0.35</td>
</tr>
<tr>
<td>Yes</td>
<td>32</td>
<td>2.16 ± 0.35*</td>
<td>1.78 ± 0.41</td>
<td>1.25 ± 0.24</td>
</tr>
<tr>
<td>Chemotherapy only</td>
<td>4</td>
<td>2.12 ± 0.52</td>
<td>1.82 ± 0.31</td>
<td>1.38 ± 0.32</td>
</tr>
<tr>
<td>Chemotherapy then radiation therapy†</td>
<td>18</td>
<td>2.13 ± 0.34‡</td>
<td>1.65 ± 0.32‡</td>
<td>1.33 ± 0.25</td>
</tr>
<tr>
<td>Focal radiation therapy†</td>
<td>2</td>
<td>2.33 ± 0.31</td>
<td>1.89 ± 0.41</td>
<td>1.24 ± 0.11</td>
</tr>
<tr>
<td>Whole, whole + focal</td>
<td>14</td>
<td>2.07 ± 0.32</td>
<td>1.59 ± 0.33</td>
<td>1.34 ± 0.27</td>
</tr>
<tr>
<td>Radiation therapy then chemotherapy‡</td>
<td>10</td>
<td>2.28 ± 0.33</td>
<td>2.01 ± 0.44</td>
<td>1.14 ± 0.15</td>
</tr>
<tr>
<td>Focal</td>
<td>2</td>
<td>2.20 ± 0.37</td>
<td>2.19 ± 0.24</td>
<td>1.00 ± 0.06</td>
</tr>
<tr>
<td>Whole, whole + focal</td>
<td>7</td>
<td>2.23 ± 0.31</td>
<td>1.86 ± 0.42</td>
<td>1.20 ± 0.15</td>
</tr>
<tr>
<td>Radiation therapy</td>
<td>No</td>
<td>26</td>
<td>2.35 ± 0.60</td>
<td>1.77 ± 0.34</td>
</tr>
<tr>
<td>Yes†</td>
<td>54</td>
<td>2.30 ± 0.40</td>
<td>1.82 ± 0.39</td>
<td>1.30 ± 0.24</td>
</tr>
<tr>
<td>Focal</td>
<td>15</td>
<td>2.48 ± 0.45</td>
<td>1.98 ± 0.36</td>
<td>1.30 ± 0.29</td>
</tr>
<tr>
<td>Whole brain</td>
<td>33</td>
<td>2.19 ± 0.35‡</td>
<td>1.71 ± 0.37‡</td>
<td>1.31 ± 0.23</td>
</tr>
<tr>
<td>Surgical resection</td>
<td>No</td>
<td>17</td>
<td>2.27 ± 0.41</td>
<td>1.74 ± 0.43</td>
</tr>
<tr>
<td>Yes</td>
<td>63</td>
<td>2.33 ± 0.49</td>
<td>1.82 ± 0.36</td>
<td>1.30 ± 0.24</td>
</tr>
</tbody>
</table>

* All averaged treatment scores were significantly different (t-test, \( P < .001 \)) when comparing the presence or absence of a treatment method.
† Details of radiation therapy portal and dose were not available for all children.
‡ Significantly different from control subjects (Tukey, \( P < .05 \)).
a variety of neural cells, within axons in both central and peripheral areas, and in nonneuronal as well as neuronal components in both white and gray matter (23, 30 –32). Tedeschi et al (33) found larger NAA signals from the centrum semiovale than from cortical or thalamic gray matter, perhaps because of contributions from other N-acetyl compounds within myelinated tissue to the NAA spectral peak. In vitro identification of NAA was reported by Urenjak et al (32), with the unexpected finding of large amounts of NAA in oligodendrocyte type 2 astrocyte progenitor cells. Their data suggested that individual cell types showed characteristic patterns detectable by proton MR spectroscopy and that these patterns could aid in identifying tumor types.

NAA may play a variety of metabolic roles associated with neuronal protein synthesis, myelination, or brain neurotransmitter activity (30). A decrease in NAA has been linked to loss of neuronal activity in demyelinating disease that may precede detectable signal intensity changes on MR images (34). Reduced NAA has also been described in response to global hypoxic-ischemic injury that is poorly characterized by MR imaging in children who have nearly drowned (16). Thus, the decrease in NAA-containing ratios identified in our frontal lobe voxels could reflect subtle damage.

The Cr peaks in a spectrum are composed of creatine and phosphocreatine. Cr is about 20% higher in concentration in gray matter than in white matter, whereas in white matter, Cho peaks are only slightly higher than in gray matter (24, 31). Ross et al (24) found that the most apparent gray-white matter MR spectroscopic difference is the higher Cho/Cr ratio in gray matter, 0.83 versus 0.59 in white matter. Since in vivo MR spectroscopic studies are limited by use of relatively large voxels that include both gray and white matter, more studies may be needed to further clarify the contribution of Cr and Cho to MR spectroscopic ratios. Cr is reported to be diminished in hypoxia, stroke, tumor, and in young infants, but also varies in response to a variety of systemic metabolic states related to hepatic and renal metabolism (24).

The Cho peak that is detectable at MR spectroscopy is composed of glycerolphosphorylcholine, phosphorylcholine, and choline (24). Increases in Cho have been associated with disease conditions involving cellular destruction with loss of membrane and/or myelin integrity (24) and with brain tumors in which increased membrane synthesis and cellularity are present (35). Phosphatidylcholine in intact membranes and myelinated tissues probably does not contribute to the Cho MR spectroscopic signal (24). Elevated Cho and lactate have been described in areas of active demyelination, although the primary effect of demyelination is reflected in diminished NAA related to neuronal and axonal injury (24). Radiation effects include loss of NAA and increase in Cho (24).

Lactate is not generally detectable in normal brain tissue by MR spectroscopy when using a small voxel size and a minimum number of transients. Lactate peaks are visible in MR spectra of tumor cysts, necrotic tissues, secondary nonneuronal tumors, and in a variety of hypoxic events (24). Although it was anticipated that we would encounter MR spectroscopic evidence of lactate, the complete absence of lactate spectral peaks from nontumorous brain tissue in this study was not entirely surprising. Lactate clears rapidly from well-perfused tissue, such as nontumorous brain. Because our voxel position was intention-

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**TABLE 4: MR spectroscopic ratios for each tumor type**

<table>
<thead>
<tr>
<th>Tumor Type</th>
<th>No. of Patients</th>
<th>NAA/Cr</th>
<th>NAA/Cho</th>
<th>Cho/Cr</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control subjects</td>
<td>11</td>
<td>2.77 ± 0.59</td>
<td>2.10 ± 0.43</td>
<td>1.34 ± 0.26</td>
</tr>
<tr>
<td>Primitive neuroectodermal</td>
<td>28</td>
<td>2.22 ± 0.36*</td>
<td>1.80 ± 0.44</td>
<td>1.29 ± 0.27</td>
</tr>
<tr>
<td>tumor</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Low grade astrocytoma</td>
<td>33</td>
<td>2.43 ± 0.55</td>
<td>1.86 ± 0.32</td>
<td>1.32 ± 0.22</td>
</tr>
<tr>
<td>Malignant astrocytoma</td>
<td>3</td>
<td>2.22 ± 0.47</td>
<td>1.66 ± 0.46</td>
<td>1.38 ± 0.37</td>
</tr>
<tr>
<td>Ependymoma</td>
<td>8</td>
<td>2.08 ± 0.56</td>
<td>1.67 ± 0.41</td>
<td>1.24 ± 0.29</td>
</tr>
<tr>
<td>Germ cell</td>
<td>4</td>
<td>2.41 ± 0.32</td>
<td>1.80 ± 0.16</td>
<td>1.35 ± 0.19</td>
</tr>
<tr>
<td>Craniopharyngioma</td>
<td>2</td>
<td>2.50 ± 0.28</td>
<td>1.49 ± 0.17</td>
<td>1.68 ± 0.00</td>
</tr>
<tr>
<td>Other</td>
<td>2</td>
<td>2.30 ± 0.14</td>
<td>1.86 ± 0.25</td>
<td>1.25 ± 0.24</td>
</tr>
</tbody>
</table>

* NAA/Cr ratios were significantly lower in these patients than in control subjects (Tukey analysis, P ≤ .05).

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Fig 3. Graphs show the relationship between NAA/Cr and whole-brain radiation in patients treated with radiation only ($r = -0.67, P < .05, n = 11$ patients) (A) and in those treated with radiation and chemotherapy ($r = .24, n = 22$ patients) (B).
alized a comparative study of $^{31}$P MR spectroscopy therapy and/or chemotherapy. Szigety et al (37) reported a comparative study of $^{31}$P MR spectroscopy (13 patients) and proton MR spectroscopy (10 patients) in radiated nontumorous adult human brain tissue adjacent to a region of neoplasia and found no detectable differences in phosphorus in the radiated tissue. Proton spectroscopy, however, revealed metabolic abnormalities that were most noteworthy in brain regions that received the highest doses of radiation. Because both Cr/Cho and NAA/Cho ratios decreased after radiation therapy, these authors postulated that the primary effect on radiated nontumorous brain tissue was a release of membrane-bound Cho related to membrane lipid breakdown at a cellular level, although it was recognized that a decrease in NAA could be contributory to the alterations in the MR spectroscopic ratio. In another study of radiation effects on nontumorous brain tissue, Usenius et al (38) reported quantitative proton MR spectroscopic findings from tissue adjacent to tumor sites in eight adults who had undergone radiation therapy. By using brain water concentration as an internal reference, these investigators found it apparent that the primary metabolic abnormality was a decrease in NAA, with relative stability of Cho and Cr values. Furthermore, Yousem et al (25) reported a reduction in NAA/Cho and NAA/Cr in brains of irradiated cats from tissue that was normal in intensity at MR imaging. The voxel position in our study was more remote from primary tumor sites than in the above studies and the brain tissue evaluated was more normal in intensity at MR imaging. The fact that the MR spectroscopic findings of declining NAA-containing ratios were greater for whole-brain radiation than for irradiation of the tumor only (Table 3) in our study suggests that radiation therapy, although not the sole explanation, may contribute to the MR spectroscopic ratio changes found in our population.

We found significantly lower NAA/Cr ratios in children who received chemotherapy as part of their treatment than in those who did not. Although most of the patients receiving chemotherapy also received radiation, patients administered chemotherapy before radiation therapy had a trend toward lower NAA/Cr and NAA/Cho (in particular), which may reflect synergistic or sensitizing effects of some chemotherapeutic agents. Moreover, although the group is small, those patients receiving only chemotherapy also had reduced NAA/Cho ratios without the lowering in NAA/Cho seen with radiation. The modulating effect of chemotherapy can also be appreciated by comparing NAA/Cr values with radiation dose. When chemotherapy was absent, there was a linear decrease in NAA/Cr with increasing dose (Fig 3A). When chemotherapy was present, the NAA/Cr values were decreased without regard to radiation dose (Fig 3B).

In this study, the MR spectroscopic data were acquired after all clinically relevant imaging sequences were completed. As a result, all patient MR spectroscopic data were acquired after administration of a paramagnetic contrast agent. While we would have preferred to acquire the MR spectroscopic data before contrast administration, doing so might have jeopardized the ability to complete the clinical study without the use of additional sedation. And although contrast material might affect MR spectroscopic ratios from tumor sites, our comparison of pre- and postcontrast patient data did not show a significant difference in MR spectroscopic ratios from nontumorous brain voxels.

For future investigations of associated effects of tumor treatment, the techniques of short-echo single-voxel MR spectroscopy and chemical shift imaging (CSI) appear promising. Short-echo MR spectroscopic studies offer a way to map more cerebral metabolites, such as amino acids, that may be more sensitive markers for the effects of brain tumor therapy. CSI offers a comparative analysis of metabolite peaks or metabolite mapping across a relatively large tissue volume (39). CSI techniques thus could be used to confirm single-voxel findings as treatment-associated effects. With further experience and larger patient populations, more subtle effects of individual treatment regimens and the relationship between treatment, MR spectroscopic findings, and clinical and neuropsychological effects could be elucidated.

**Conclusions**

Our findings indicate that the effects of brain tumor therapy are associated with measurable changes in NAA/Cho and NAA/Cr ratios from nontumorous brain tissue in children with primary brain neoplasms as compared with healthy children. The most consistent finding was a decrease in NAA/Cr. Chemotherapy as one component of a multitreatment protocol was associated with significant reduction in the NAA/Cr ratio. There was a trend for patients treated with chemotherapy before radiation therapy to have lower NAA/Cr and NAA/Cho ratios, suggesting that chemotherapy may alter the subsequent biochemical effects of radiation therapy. Although we anticipated a strong relationship between the MR spectroscopic ratios and the use of radiation therapy, we instead found a trend toward lower NAA/Cr and NAA/Cho ratios in children treated with whole-brain radiation relative to those treated only with focal tumor therapy and to those not treated with radiation therapy. Although further experience is needed, our study suggests a potential role for MR spectroscopy in moni-
toring the effects of treatment on nontumorous brain tissue.

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