

ScholarWorks@GSU

Estrogen Receptor Alpha in the Medial Preoptic Area Mediates Male Rat Sexual Responses to Estrogen

Authors	Russell, Nancy
Citation	Russell, Nancy. (2010). Estrogen Receptor Alpha in the Medial Preoptic Area Mediates Male Rat Sexual Responses to Estrogen. Georgia State University. https://doi.org/10.57709/1397246
DOI	https://doi.org/10.57709/1397246
Download date	2026-05-08 11:00:35
Link to Item	https://hdl.handle.net/20.500.14694/2271

ESTROGEN RECEPTOR ALPHA IN THE MEDIAL PREOPTIC AREA MEDIATES MALE
RAT SEXUAL RESPONSES TO ESTROGEN

by

NANCY RUSSELL

Under the Direction of Andrew N. Clancy, PhD

ABSTRACT

Male rat sexual behavior requires aromatization of testosterone (T) to estradiol (E₂) in the medial preoptic area (MPO) where estrogen receptors (ER) exist in two isoforms, ER α and ER β . We hypothesized that E₂ acts through estrogen receptor α (ER α) in the MPO to promote male mating behavior. Four groups of male rats were castrated, administered DHT s.c. and bilateral MPO implants delivering either: cholesterol, E₂, propyl pyrazole triol (PPT, ER α agonist), diarylpropionitrile (DPN, ER β agonist), or 1-methyl-4-phenyl pyridinium (MPP, ER α antagonist). Additional gonadally intact males received bilateral MPO DPN implants. PPT maintained sexual behavior equally as well as E₂, whereas mating was not maintained by cholesterol or DPN MPO implants. Exogenous T did not reinstate mating in animals that received MPP MPO implants. These findings indicate that, in the MPO, ER α is necessary and sufficient to promote copulatory behavior in male rats and ER β is not sufficient for mating.

INDEX WORDS: Medial preoptic area, Estrogen receptor, Estradiol, Dihydrotestosterone, propyl-pyrazole-triol (PPT), Cholesterol, Sexual behavior, Diarylpropionitrile (DPN), 1-methyl-4-phenyl pyridinium (MPP)

ESTROGEN RECEPTOR ALPHA IN THE MEDIAL PREOPIC AREA MEDIATES MALE
RAT SEXUAL RESPONSES TO ESTROGEN

by

NANCY RUSSELL

A Thesis Submitted in Partial Fulfillment of the Requirements for the Degree of

Master of Science

in the College of Arts and Sciences

Georgia State University

2010

Copyright by
Nancy Russell
2010

ESTROGEN RECEPTOR ALPHA IN THE MEDIAL PREOPTIC AREA MEDIATES MALE
RAT SEXUAL RESPONSES TO ESTROGEN

by

NANCY RUSSELL

Committee Chair: Andrew N. Clancy

Committee: Matthew Grober

Kyle Frantz

Electronic Version Approved:

Office of Graduate Studies

College of Arts and Sciences

Georgia State University

August 2010

ACKNOWLEDGMENTS

I am extremely grateful to have Dr. Andrew Clancy as my thesis advisor and for providing the opportunity to further my education. I also want to thank him for his leadership and team spirit during my training. I also express my appreciation to the members of my committee, Dr. Kyle Frantz and Dr. Matthew Grober, for their time and effort. I thank Carrie Paisley for instruction and for her friendship. I thank LaTasha Warren-Morrison for always being there when I needed it and for helping out with graduate school. I am also grateful for to everyone that has been a part of this journey being successful. Dr. Sarah Pallas thank you for always pushing and requiring nothing less than all we are capable of, even when it was petrifying. Lastly, I would like to thank my entire family, especially my sister Tania Ettema for her resilience and determination; I have borrowed it many times. I would like to thank my parents Judy Geisinger and Johnny Russell. Kelli Eastman, thank you for patiently listening to everything.

TABLE OF CONTENTS

ACKNOWLEDGEMENTS	iv
LIST OF TABLES	vi
LIST OF FIGURES	vii
INTRODUCTION	1
MATERIALS AND METHODS	4
<i>Animals</i>	3
<i>Female rat surgical procedures</i>	4
<i>Sexual behavior tests</i>	4
<i>Experiment 1</i>	5
<i>Male rat experimental groups and surgery</i>	5
<i>Experiment 2 procedures</i>	6
<i>Male experimental groups and surgery</i>	6
<i>Histological Verification</i>	7
<i>Statistical analyses</i>	7
RESULTS	8
<i>Experiment 1</i>	8
<i>Experiment 2</i>	11
DISCUSSION	12
REFERENCES	17

LIST OF TABLES

TABLE 1: TABLE 1.1 Statistical Analysis MPP Experiment

12

LIST OF FIGURES

Figure 1: Male Sexual Behavior	32
Figure 2: Experiment 1 Cannula Placements MPO	33
Figure 3: Experiment 2 Cannula Placements MPO	34

INTRODUCTION

Male rat mating behavior is dependent on testosterone (T), which is enzymatically converted in the brain into estradiol (E₂) by aromatase [Naftolin et al., 1975] and into the non-aromatizable androgen, dihydrotestosterone (DHT) by 5 α reductase [Martini, 1982; Massa et al., 1972]. These metabolites, together with T, act on steroid sensitive neurons, including those containing estrogen receptors (ER) in the medial preoptic area (MPO) of the male rat brain to promote mating behavior [Vagell and McGinnis, 1997]. Thus, mating declines after castration and is reinstated by exogenous T [Christensen and Clemens, 1974; Davidson, 1966a, 1966b, 1996]. Furthermore, castrated male rats mate normally (to ejaculation) after being treated with a combination of E₂ and DHT, given in physiological dose ranges, and their mating behavior is comparable to that of rats treated with physiological levels of T [Baum and Vreeburg, 1973; Larsson et al., 1973; Davidson, 1966a, 1966b, 1996]. When administered separately, however, neither E₂ [Baum and Vreeburg, 1973; Davidson, 1969] nor DHT [Feder, 1971; McGinnis and Dreifuss, 1989] fully maintains mating behavior, indicating that estrogens and androgens both are necessary for the normal expression of male rat sexual behavior. Fadrozole, a non-steroidal aromatase inhibitor [Lipton et al., 1990; McGinnis and Mirth, 1986; McGinnis et al., 1996; Vagell and McGinnis, 1997] that blocks formation of E₂ from T, administered either systemically [Bonsall et al., 1992] or intracerebrally into the MPO [Clancy et al., 1995] attenuates male rat mating behavior, an effect that can be partially reversed via administration of E₂ [Bonsall et al., 1992]. Mating is also maintained by implants of E₂ into the MPO of gonadally intact male rats receiving fadrozole s.c. that blocks E₂ formation elsewhere throughout the body and brain [Clancy et al., 2000].

Studies of mating induced expression of Fos immunoreactivity (ir) show that induced Fos-ir occurs in MPO cells that contain receptors for gonadal steroids, including those that express ER-ir [Baum and Everitt, 1992; Coolen et al., 1996; Gréco et al., 1996; Greco et al., 1998; Gréco et al., 2003]. Steroid sensitive cells of the MPO are critical for expression of male rat mating behavior [Brackett and Edwards, 1984; Larsson and Heimer, 1964; Simerly and Swanson, 1986].

Multiple subtypes of ER exist [Greco et al., 2003 ; Kuiper et al., 1996], some of which may be located within the cell membranes of E₂ responsive neurons [Evinger and Levin, 2005; Thomas et al., 2005]. The second type of ER to be discovered was termed ER β and the original ER is now known as ER α [Kuiper et al., 1996]. ER α and ER β are both expressed in steroid sensitive neurons of the rat MPO [Abraham et al., 2004; Greco et al., 2003; Nomura et al., 2003, Shughrue and Merchenthaler, 2001]. Evidence from receptor knockout experiments in male mice suggests that ER α may play a dominant role, and ER β a minor role, in the expression of male sexual behavior [Ogawa et al., 1997; Ogawa et al., 1998; Ogawa et al., 2000; Rissman et al., 1997]. Also, in male rats an anti-sense oligodeoxynucleotide (AS-ODN), complimentary to ER α mRNA, reduced the expression of ER α -ir when infused bilaterally into the MPO of gonadally intact male rats, which in turn resulted in decreased mating behavior [Paisley, 2007; Paisley et al., 2005]. This suggests that, in the MPO, estrogenic metabolites of T act via ER α , and that ER α is necessary in the MPO for male rat mating behavior to occur.

The recent development of highly selective estrogenic drugs that modulate ER α and/or ER β function as powerful tools that may aid in identifying the behaviorally relevant ER isoform in the male rat MPO for sexual responses to E₂ [Harrington et al., 2003; Hillisch et al., 2004]. Therefore, in the present study, we used selective estrogenic drugs implanted into the MPO to

test the hypothesis that, in the MPO, ER α mediates sexual responses of male rats to E $_2$. In two experiments, proven ejaculators were divided into matched groups based on total ejaculation frequency during presurgical sexual behavior tests. In the first experiment, four groups of male rats were castrated and administered DHT s.c. and then received bilateral MPO implants carrying one of the following drugs: i) cholesterol (negative control), ii) E $_2$ (positive control), iii) propyl pyrazole triol (PPT, an ER α agonist) or iv) diarylpropionitrile (DPN, an ER β agonist). A fifth group was used as a toxicity control that consisted of gonadally intact male rats that also received bilateral MPO DPN implants. In the second experiment, two groups of male rats were castrated, tested until mating behavior declined, and then implanted with T s.c. and bilateral MPO implants delivering either no drugs (blank) or the drug 1-methyl-4-phenyl pyridinium (MPP, an ER α antagonist) [Zhou et al., 2009].

For the first experiment, we predicted that PPT, the ER α agonist, implanted into the MPO would mimic the action of E $_2$ and fully maintain the mating behavior of castrated, DHT maintained male rats. On the other hand, DPN, an ER β agonist, would not support mating behavior following its implantation into the MPO. Moreover, we predicted that MPO implants of MPP, an ER α antagonist, would block T-induced reinstatement of mating in castrated male rats.

MATERIALS AND METHODS

Animals

Sprague Dawley male and female rats, obtained from Charles River, were given free access to laboratory chow and tap water and kept in polycarbonate shoebox, 22 x 44 x 18 cm, at Georgia State University. The rats were acclimated to a 14:10 hour reverse light: dark cycle (lights on at 1900 EST). Male rats were housed two per cage until surgery, after which they were housed individually. Females remained paired throughout the study. Animal care was in accordance with humane standards (NIH publ. No. 85-23, revised 1985) and all procedures involving animals were authorized by the Georgia State University IACUC.

Female rat surgical procedures

Females were anesthetized with isoflurane gas in a chamber (5% gas at 1.0 liters per minute oxygen exchange rate) and transferred to a nosecone (2-3% gas at 0.3 liters per minute oxygen). Ovaries were removed through an abdominal incision and a Silastic capsule (6mm length, 1.981 mm i.d. x 3.175 mm o.d.) containing crystalline E₂ was implanted s.c. into the scapular region. Females were allowed at least five days to recover from surgery before being paired with male rats to measure sexual behavior in the males. Four hours prior to each weekly behavioral test, females were injected s.c. with 1 mg progesterone in 0.2 ml of sesame oil to induce sexual receptivity.

Sexual behavior tests

Male rats were paired with sexually receptive females and screened once weekly for three consecutive weeks for the display of sexual behavior prior to being selected for the experiment. Tests for sexual behavior were conducted by blind observers once a week, 30 minutes per test. Sexual behavior testing took place under red light illumination during the dark phase of the light:

dark cycle. The recorded behaviors during each 30 minute test included: mount frequency (MF): number of mounts with no penetration; intromission frequency (IF): numbers of mounts with penetration; ejaculation frequency (EF): numbers of ejaculation; mount latency (ML): time from test start until the first mount or intromission or 1800 seconds if mounts did not occur; ejaculation latency (EL): time from first mount or intromission until the first ejaculation or 1800 seconds if ejaculation did not occur; post ejaculatory interval (PEI): the time between the first ejaculation and the next mount or intromission or 1800 seconds if ejaculation did not occur. The same sex behavior testing procedures were used after males had recovered from surgery.

Experiment 1

Male rat experimental groups and surgery

Upon completion of sexual screening trials, four groups of male rats, all proven ejaculators, were matched and divided into groups, according to total numbers of ejaculations during sexual screening trials, then males were castrated, administered DHT s.c., and received bilateral MPO implants carrying one of the following drugs: i) cholesterol (negative control), ii) E₂ (positive control), iii) PPT (ER α agonist) or iv) DPN (ER β agonist). A fifth matched group of gonadally intact male rats served as DPN toxicity controls; these animals were not castrated but received bilateral MPO implants of DPN that were identical to those used in the DPN group (see above iv). Specifically, males were anesthetized with isoflurane gas in a chamber and transferred to a nosecone as described above for females, castrated through midline abdominal incisions, and received s.c. in the scapular region a 10-mm Silastic capsule (1.981 mm ID x 3.175 mm OD) containing crystalline DHT. DHT capsules of these dimensions have been reported to produce circulating DHT levels in the physiological range [Ando et al., 1998; Lugg et al., 1995; Parte and Juneja, 1992]. They were then placed in a stereotaxic instrument and

implanted bilaterally with ethylene oxide sterilized 22 gauge stainless steel guide cannulae (Plastics One, Roanoke, VA, USA) aimed at the MPO (level skull coordinates with respect to bregma: anterior-posterior = -0.5 mm, medial-lateral = \pm 0.75 mm, dorsal-ventral = -8.0 mm [Swanson, 1998]. Males serving as gonadally intact DPN toxicity controls were treated identically except they were neither castrated nor given DHT. Sterile, 28 gauge stainless steel inner cannulae, extended 1 mm below the guide cannulae; these were tamped in either E₂, DPN, PPT, or cholesterol to deliver the appropriate drug into the MPO. Thereafter, rats were allowed at least five days to recover from surgery before postoperative behavioral testing began. Sterile inner cannulae were replaced 48 hours before and 24 hours after each behavior test under brief isoflurane anesthesia. Tips of cannulae were examined microscopically after they were removed from the brain to ensure drug was present, none failed.

Experiment 2

Male rat experimental groups and surgery

MPP (ER α antagonist) or blank (control) drug carrying MPO cannulae were implanted in castrated male rats that were administered T exogenously to test if MPP would interfere with T-induced reinstatement of mating. Specifically, male rats were screened for sexual behavior and proven ejaculators were matched on total numbers of ejaculations. Thereafter the males were anesthetized as described previously and castrated through midline abdominal incisions. After a recovery period of at least five days, the castrated males were tested with receptive females once a week until all mating ceased. Each male was then anesthetized as described before and implanted s.c. with a 15 mm Silastic capsule (1.981 mm ID x 3.175 mm OD) containing crystalline T. T capsules of this size have been reported to produce circulating T levels in the physiological range [Verjans et.al., 1975]. Immediately thereafter, they were stereotaxically

implanted bilaterally with 22-gauge stainless-steel, ethylene oxide-sterilized, guide cannulae (Plastics One, Roanoke, VA, USA) aimed at the MPO (level skull coordinates with respect to bregma: anterior-posterior = -0.5 mm, medial-lateral = ± 0.75 mm, dorsal-ventral = -8.0 mm [Swanson, 1998]. Sterile stainless steel inner cannulae (28 gauge, Plastics One) extended 1 mm below the guide cannulae and were tamped in either: (i) MPP or (ii) nothing (blank). Males were given 5–7 days to recover before the weekly postoperative behavioral testing resumed. Freshly tamped sterilized inner cannulae were inserted, under brief isoflurane anesthesia, into the brain 48 h before and 24 h after every behavioral test to ensure continuous drug delivery to the brain. Tips of cannulae were examined microscopically after they were removed from the brain to ensure drug was present, none failed.

Histological Verification

When the testing period was concluded, males were euthanized with an overdose of sodium pentobarbital (1.0 mg/kg i.p.) and perfused transcardially for 5 min with physiological saline followed by a minimum of 300 ml of a fixative containing 2% paraformaldehyde and 0.1% picric acid in 0.1 M phosphate buffer (PB) solution (pH 7.4). Skulls were partially opened and immersed overnight in perfusion fixative. Brains were removed from skulls the next day and stored in 30% sucrose in 0.1 M PB for approximately 48 h. Frozen coronal brain sections (40 μm) through the diencephalon were collected into 0.1 M PB, mounted on clean gelatin albumin-coated slides, stained with toluidine blue and examined microscopically to verify cannula placement.

Statistical analysis

In the first experiment, between-group behavioral data were analyzed by repeated measures (group by trial) analysis of variance for the pre-operative trials (5 groups by 3 trials)

and for the post operative trials (5 groups by 4 trials), followed by *post hoc* comparisons using the Tukey honestly significant differences test at a probability level of 0.05 [Kirk, 1968].

Within-group comparisons of preoperative and postoperative changes in behavior were then analyzed by paired t-tests [Kirk, 1968]. In the second experiment, group differences were analyzed by the Mann-Whitney U test [Siegel, 1956]. Data from all males were used in the statistical analyses; default values (see sexual behavior tests) were used in situations where animals did not display a given behavior. Two-tailed probabilities are reported in all cases.

RESULTS

Experiment 1

Male sexual behavior

The mating behavior of sexually experienced, castrated male rats receiving DHT s.c. was maintained by either E₂ or PPT implants to the MPO, but neither cholesterol nor DPN MPO implants maintained mating (FIGURE 1). Gonadally intact DPN toxicity controls that received DPN MPO implants also mated robustly. Specifically, the two groups receiving either E₂ or PPT MPO implants and gonadally intact DPN toxicity controls mated vigorously on all sexual behavior tests throughout the study and these three groups were statistically indistinguishable from each other on all measures of copulatory behavior during both the preoperative and postoperative periods; sexual behavior did not change significantly within these three groups before vs. after treatment. In contrast, sexually experienced, castrated, DHT s.c. treated males that received either cholesterol or DPN implants into the MPO virtually ceased mating during the postoperative period whereas they had mated robustly prior to surgery. It is noteworthy that, during the preoperative period, all five groups mated vigorously and no group differences were

observed. Thus, MPO implants of E₂ itself or PPT (ER α agonist) but not cholesterol or DPN (ER β agonist) maintained mating behavior.

There were no significant group differences on any of the six measures of copulatory behavior during the preoperative trials, nor were there any significant interactions. Significant behavioral differences emerged, however, during the postoperative period (FIGURE 1). Specifically, during the postoperative period, the groups differed significantly from each other on mount frequency ($F_{4,36} = 11.42$, $P < 0.001$), ejaculation frequency ($F_{4,36} = 9.79$, $P < 0.001$), intromission frequency ($F_{4,36} = 19.31$, $P < 0.001$), mount latency ($F_{4,36} = 13.09$, $P < 0.001$), ejaculation latency ($F_{4,36} = 5.52$, $P < 0.002$), and the post-ejaculatory interval ($F_{4,36} = 9.23$, $P < 0.001$). No other statistically significant main effects or interactions were observed during the postoperative period. Follow-up post-hoc analysis of sexual behavior during the postoperative period revealed no significant differences on any test day between the E₂ group compared to either the PPT or gonadally intact DPN toxicity control groups on any of the six behavioral measures all of which groups continued to mate vigorously, except that, on the first postoperative trial, rats receiving MPO E₂ implants ejaculated significantly more frequently than those that received MPO PPT implants ($P < 0.05$). In contrast, the E₂ group mated significantly ($P < 0.05$) more than the cholesterol group on all postoperative test days and on all six behavioral measures except mount frequency on the first postoperative trial, ejaculation frequency on the fourth postoperative trial, ejaculation latency on the first and fourth postoperative trials and the post-ejaculatory interval on the first postoperative trial. Similarly, the E₂ group mated significantly ($P < 0.05$) more than the DPN group on all postoperative test days and on all six behavioral measures except mount frequency on the first and fourth postoperative trials and mount latency, ejaculation latency and the post-ejaculatory interval on the first postoperative trial. In addition,

the cholesterol group mated significantly ($P < 0.05$) less than the PPT group on all postoperative test days and on all six behavioral measures except mount frequency, ejaculation frequency and ejaculation latency on the first and fourth postoperative trials and the post-ejaculatory interval on the first post operative trial, whereas the cholesterol group mated significantly ($P < 0.05$) less than the gonadally intact DPN toxicity control group on all postoperative test days and on all six behavioral measures except ejaculation frequency on the second postoperative trial, ejaculation latency on the first, second and fourth postoperative trials and the post-ejaculatory interval on the first postoperative trial. Finally, the cholesterol group differed significantly ($P < 0.05$) from the DPN group only on ejaculation latency on the fourth postoperative trial.

Changes in sexual behavior within each group were examined by comparing the performances on the last preoperative testing day and the last postoperative testing day. There were no significant changes on any of the six indices of copulatory behavior in gonadally intact DPN toxicity controls, nor did the groups of castrated males maintained on DHT s.c. that received MPO implants of either E_2 or PPT differ significantly on any behavioral measure before and after surgery. However, compared to their preoperative mating performances, sexual behavior deteriorated significantly during the postoperative period in the groups of castrated males maintained on DHT s.c. and implanted in the MPO with either cholesterol or DPN. Specifically, in the cholesterol group, mount frequency ($t = 6.21$, $P < 0.001$) and intromission frequency ($t = 10.32$, $P < 0.001$) were significantly depressed on the last postoperative test compared with the last preoperative day (FIGURE 1), whereas mount latency ($t = 3.95$, $P < 0.003$) and the post-ejaculatory interval ($t = 3.03$, $P < 0.01$) were significantly longer on the last postoperative test relative to the last preoperative day. Similarly, in castrated, DHT s.c.-maintained male rats receiving DPN MPO implants, ejaculation frequency ($t = 4.67$, $P < 0.003$)

and intromission frequency ($t = 5.73, P < 0.001$) were significantly depressed on the last postoperative test compared with the last preoperative day (FIGURE 1), whereas mount latency ($t = 3.86, P < 0.008$), ejaculation latency ($t = 5.86, P < 0.001$) and the post-ejaculatory interval ($t = 3.82, P < 0.009$) were significantly longer on the last postoperative test relative to the last preoperative day.

Cannulae Placements

Locations of cannula tips are shown in FIGURE 2. Cannula tips were located in or near the MPO or anterior hypothalamus for all males

Experiment 2

Male sexual behavior

Exogenous T rapidly reinstated mating behavior in control males that received blank MPO implants. In contrast, exogenous T did not promote significant restoration of copulatory behavior in animals that received MPP MPO implants, indicating MPP interfered with T-induced reinstatement of sexual behavior. Specifically, on trial 2, rats receiving blank MPO implants mounted ($U_{(n1=5, n2=4)} = 0, p < 0.014$) and intromitted ($U_{(n1=5, n2=4)} = 0, p < 0.014$) significantly more frequently than animals that received MPP MPO implants and, on trial 2, there was a non-significant trend toward greater ejaculation frequency in the blank MPO groups than in the MPP MPO group ($U_{(n1=5, n2=4)} = 2.5, p < 0.066 = ns$), TABLE 1. It is noteworthy that, by the second trial, all males that had received blank MPO implants mounted and intromitted and three of these four rats mated to ejaculation. In contrast only one of the five males in the MPP MPO group mounted and none intromitted or ejaculated. Mount latency, Ejaculation Latency and PEI data are not shown.

Cannulae Placements

Locations of cannula tips are shown in FIGURE 3. Cannula tips were located in the MPO or anterior hypothalamus for all males.

TABLE 1.1 Statistical Analysis MPP Experiment

GROUP:	BLANK to MPO (N=4)		MPP to MPO (N=5)	
BEHAVIOR:	Trial 1	Trial 2	Trial 1	Trial 2
Mount Frequency	15.25 ± 8.65	*22.25 ± 3.59 *p < 0.014 *Blank vs MPP, Trial 2 MF	1.2 ± 1.2	*0.2 ± 0.2
Intromission Frequency	11.75 ± 7.15	*14.25 ± 5.02 *p < 0.014 *Blank vs MPP, Trial 2 IF	0 ± 0	*0 ± 0
Ejaculation Frequency	0.75 ± 0.48	1.5 ± 0.5	0 ± 0	0 ± 0

DISCUSSION

The objective of these experiments was to test the hypothesis that, in the MPO, ER α mediates the sexual responses of male rats to E₂. This hypothesis was confirmed by examination of the copulatory response of animals implanted in the MPO with various selective ER α or ER β agonists or antagonists. We found that, in the MPO, ER α alone is necessary and sufficient to promote mating behavior. Specifically, the mating behavior of sexually experienced, castrated male rats receiving DHT s.c. was fully maintained by MPO implants of either E₂ or PPT (ER α agonist) but neither MPO implants of DPN (ER β agonist) nor cholesterol were effective in maintaining copulatory behavior. Because the group of gonadally intact males with DPN MPO implants (DPN toxicity controls) mated vigorously on all sexual behavior tests before and after

surgery and did not differ significantly from the two groups receiving either E₂ or PPT MPO implants, it is unlikely that DPN is toxic to the brain, nor does it interfere with mating. Moreover, MPO implants of MPP (ER α antagonist) inhibited T-induced reinstatement of sexual behavior, which extends and replicates, by an independent method, the findings in a companion report that blocking the synthesis and expression of ER α in the MPO reduces sexual behavior [Paisley, 2007; Paisley et al., 2005]. It is unlikely that MPP is toxic to the brain or interferes non-specifically with mating because in other experiments in our laboratory [Ogaga-Mgbonyebe et al., 2009], MPP was implanted into the amygdala of gonadally intact male rats, which animals continued to mate vigorously. This pattern of results indicates that, in the MPO, ER α is critical for mating. Taken together these findings support the interpretation that, in the MPO, ER α exclusively mediates sexual responses to E₂ and, in this brain area, ER α is necessary and sufficient to promote mating behavior in male rats, whereas, in the MPO, ER β is not sufficient for the expression of this behavior.

In this study we report on the effects on male rats' copulatory behavior of the selective ER agonists PPT and DPN and antagonist MPP following their direct, site-specific implantation to the brain. Previous studies have reported only in female rats, that received intraventricular application of PPT but not DPN, which promoted lordosis, presumably by action on the ventromedial hypothalamus [Mazzuco et al., 2008; Walf and Frye, 2005; Walf et al., 2007]. In male rats, steroid sensitive MPO neurons expressing ER alone, or androgen receptors (AR) only, or that co-express both AR and ER are activated by mating [Greco et al., 1998;]. However, ER exists in at least two isoforms, ER α and ER β [Kuiper et al., 1996]. Knockout (KO) studies in male mice suggest that the form of ER that is centrally important for display of male sexual behavior is ER α [Ogawa et al., 1997; Ogawa et al., 1998; Ogawa et al., 2000; Rissman et al.,

1997]. Nevertheless, whereas ER α KO profoundly reduced mating [Ogawa et al., 1997, Ogawa et al., 1998] and ER β KO showed only a minimal reduction [Scordalakes et al., 2003], the combined knockouts of ER α and ER β [Owaga et al., 2000] virtually eliminated mating, suggesting a major role for ER α and a minor role for ER β in male mouse mating. Several other laboratories [Davis and Barfield, 1979; Greco et al., 1998; Greco et al., 2003; Walf et al., 2007] have shown that ER α or ER β mediate various reproductive and non-reproductive behaviors by their actions in the brain. In female rats, ER α -ir and ER β -ir neurons in MPO and MEA were both activated by sexual behavior and expressed mating-induced Fos-ir, raising the possibility that both ER subtypes make a contribution to sexual behavior [Greco et al., 2003]. Previous findings from our laboratory show that estrogenic metabolites of T act in the MPO to promote mating behavior in male rats [Clancy et al., 1995, 2000]. These findings were extended by experiments with gonadally intact male rats in which an anti-sense oligodeoxynucleotide (AS-ODN) complimentary to ER α mRNA was infused into the MPO, which reduced significantly the expression of ER α -ir in the MPO and resulted in significantly decreased mating behavior [Paisley, 2007; Paisley et al., 2005]. These findings, together with those in the present study, suggest ER α -related signal transduction pathways in the MPO are important in mediating male rat sexual responses to E₂ but that neither ER β /ER β homodimerization nor ER α /ER β heterodimerization in the MPO are necessary for mating. Moreover, MPO implants of either E₂ conjugated to bovine serum albumin (BSA-E₂) or unconjugated (free) E₂ were equally effective in maintaining male rat mating behavior [Huddleston et al., 2006], suggesting the possible existence in the MPO of an ER α signal transduction pathway that is mediated by a plasma membrane bound version of ER α (mER α). This is because BSA-E₂ is believed to prevent the entry of E₂ into the interior of target cells thereby confining its action to the cell surface

[Huddleston et al., 2006; Pappas et al., 1995]. Therefore, a novel mechanism of E₂ action may exist in the MPO in which an mER α influences the sexual behavioral responses to E₂. If such a mER α MPO signal transduction pathway exists, then it may be possible for the MPO to be quickly and easily activated whenever the social or environmental circumstances are ideal for mating behavior to be expressed [Abraham et al., 2004; Chaban et al., 2004; Pietras and Szego, 1977]. Stimulus-induced fluctuations in levels of circulating gonadal steroids such as female-induced or pheromonally-induced elevations in gonadotropins and T [Clancy et al., 1984, Coquelin et al., 1984, Khan et al., 2005; Macrides et al., 1975], if accompanied by rapid aromatization of T to E₂, conceivably could activate the MPO via a mER α to promote rapidly the expression of male mating behavior.

Different ligands such as PPT, DPN, MPP or E₂ bind selectively to different ER subpopulations thereby eliciting different responses [Staufer et al., 2000; Wright et al., 2006]. Cholesterol presumably is inert and does not activate either ER α or ER β , thus after cholesterol MPO implantation, mating ceased. In contrast, PPT binds ER α with a high affinity and exhibits limited cross reactivity with ER β [Stauffer et al., 2000] whereas DPN binds ER β robustly and cross reacts weakly with ER α [Katzenellenbogen et al., 2000], and E₂ activates both ER α and ER β and promotes male rat mating behavior. MPO implants of MPP, interfered with T-induced reinstatement of mating consistent with previous reports MPP is a selective ER α antagonist that has high affinity for ER α and little cross reactivity for ER β [Sun et al., 2002; Zhou et al., 2009]. These characteristics of the selective estrogenic drugs used in this study further suggest that, in the MPO, ER α mediates responses to E₂ to promote sexual behavior. Taken together, these findings indicate that, in the MPO, ER α is necessary and sufficient to mediate male rat sexual behavioral responses to E₂.

The central findings from other studies in our laboratory [Paisley, 2007; Paisley et al., 2005, Ogaga et al., 2009] indicate, however, that E_2 acts differently in the MPO than in the MEA and there may be a role for $ER\beta$ to play, outside the MPO, in the expression of male mating behavior. Thus, many questions remain how these ER subtypes promote expression of copulatory responses. For example, either $ER\alpha$ or $ER\beta$ may simply be the most numerous ER subtype in any given brain region and thus govern the overall response of that brain region to E_2 . Although previous findings [Greco et al., 1998; Greco et al., 2003] suggest it is unlikely that the different ER subtypes are each selectively associated with the expression of specific behaviors, [Toufexis et al., 2007] addressed the possibility that the ambiguous effects of E_2 in the MEA may result from mutually opposing interactions between $ER\alpha$ and $ER\beta$ at the cellular level. If this is case, then differences in the $ER\alpha/ER\beta$ ratios in different brain regions might influence the response of any given brain area to E_2 . Based on the findings in this report, we infer that, in the MPO of castrated male rats receiving DHT s.c., E_2 mediates mating via $ER\alpha$ and that this pathway alone is necessary and sufficient to activate male sexual behavior.

In conclusion, this study provides evidence suggesting that, in the MPO, $ER\alpha$ is the behaviorally relevant form of ER mediating male rat mating behavior. Thus, following MPO implantation of PPT, males mated robustly at rates that were statistically indistinguishable from those implanted in the MPO with E_2 . The effects of any ligand, such as E_2 , are mediated by the specific receptors that bind them. Once bound, there are at least two ways that E_2 may affect its targets, either by activating the traditional genomic pathway [Jensen et al., 1968; King and Greene, 1984] or via a rapid non-genomic pathway that initiates synthesis of intracellular second messengers [Minami et al., 1990; Pietras and Szego, 1977, 1980]. Further investigation is needed to determine the mechanism of action of MPO $ER\alpha$. It is noteworthy, however, that in

the MPO, BSA-E₂, which is believed not to penetrate into the interior of cells, supported vigorous mating behavior [Huddleston et al., 2006], which raises the possibility that a plasma membrane-associated ER α in neurons of the MPO mediates male rat sexual responses to E₂. From the present study, it may be concluded that, in the MPO, ER α alone is necessary and sufficient to maintain mating in castrated, androgen supplemented male rats.

REFERENCES

- Abraham, I.M., Todman, M.G., Korach, K.S., Herbison, A.E., 2004. Critical *in vivo* roles for classical estrogen receptors in rapid estrogen actions on intracellular signaling in mouse brain. *Endocrinology*. 145, 3055-3061.
- Ando S., Aquila S., Beraldi E., Canonaco M., Panno M.L., Valenti A., Dessi-Fulgheri F., 1988. Physiological changes in androgen plasma levels with elapsing of time from Castration in adult male rats. *Horm. Metab. Res.* 20, 96–99.
- Baum, M.J., Vreeburg, J.T.M., 1973. Copulation in castrated male rats following combined treatment with estradiol and dihydrotestosterone. *Science* 182(109), 283-285.
- Baum, M.J., 2003. Activational and organizational effects of estradiol on male behavioral neuroendocrine function. *Scandinavian Journal of Psychology* 44, 213-220.
- Baum, M.J., Everitt, B.J., 1992. Increased expression of c-fos in the medial preoptic area after mating in male rats: role of afferent inputs from the medial amygdala and midbrain central tegmental field. *Neuroscience* 50, 627-646.
- Bonsall, R.W., Clancy, A.N., Michael, R.P., 1992. Effects of the nonsteroidal aromatase inhibitor, fadrozole, on sexual behavior in male rats. *Horm. Beh.* 20, 240-254.

- Brackett, N.L., Edwards, D.A., 1984. Medial preoptic connections with the midbrain tegementum are essential for male sexual behavior. *Physiol. Behav.* 32, 79-84.
- Chaban, V.V., Lakhter, A.J., Micevych, P., 2004. A membrane estrogen receptor mediates intracellular calcium release in astrocytes. *Endocrinology* 145, 3788–3795.
- Christensen, L.W. and Clemens, L.G., 1975. Blockade of testosterone-induced mounting behavior in the male rat with intracranial application of the aromatization inhibitor, andros-1,4,6-triene-3,17-dione. *Endocrinology* 97, 1545-1551.
- Clancy, A.N., Coquelin, A., Macrides, F., Gorski, R.A., Noble, E.P., 1984. Sexual behavior and aggression in male mice: involvement of the vomeronasal system. *J. Neurosci.*, 4, 2222-2229.
- Clancy, A.N., Zumpe, D., Michael, R.P., 1995. Intracerebral infusion of an aromatase inhibitor, sexual behavior and brain estrogen receptor-like immunoreactivity in intact male rats. *Neuroendocrinology* 61, 98-111.
- Clancy, A.N., Zumpe, D., Michael, R.P., 2000. Estrogen in the medial preoptic area of male rats facilitates copulatory behavior. *Horm. Behav.* 38, 86-93.

- Coolen, L.M., Peters, H.J.P.W., Veening, J.G., 1996. Fos immunoreactivity in the rat brain following consummatory elements of sexual behavior: a sex comparison. *Brain. Res.* 738, 67-82.
- Coquelin, A., Clancy, A.N., Macrides, F., Noble, E.P, Gorski, R.A., 1984. Pheromonally induced release of luteinizing hormone in male mice: involvement of the vomeronasal system. *J. Neurosci.*, 4, 2230-2236.
- Davidson, J.M., 1966a. Activation of the male rat's sexual behavior by intracerebral implantation of androgen. *Endocrinology* 79, 783-794.
- Davidson, J.M., 1966b. Characteristics of sexual behaviour in male rats following castration following combined treatment with estradiol and dihydrotestosterone. *Anim. Behav.* 14, 266-272.
- Davidson, J.M., 1969. Effects of estrogen on the sexual behavior of male rats. *Endocrinology* 84, 1365-1372.
- Davis, P.G., Barfield, R.J., 1979. Activation of sexual behavior by intracranial estradiol benzoate implants in male rats. *Neuroendocrinology* 28, 217-227.
- Evinger, III A.J., Levin, E.R., 2005. Requirements for estrogen receptor α membrane localization and function. *Steroids* 70, 361-363.

- Feder, H.H., Naftolin, F., Ryan, K.J., 1974. Male and female sexual responses in male rats given estradiol benzoate and 5 alpha-androstan-17 beta-ol-3-one propionate. *Endocrinology* 94, 136-141.
- Gréco, B., Edwards, D.A., Michael, R.P., Clancy, A.N., 1996. Androgen receptor immunoreactivity and mating induced fos expression in forebrain and midbrain structures in the male rat. *Neuroscience* 75, 161-171.
- Gréco, B., Edwards, D.A., Michael, R.P., Clancy, A.N., 1998. Androgen receptors and estrogen receptors are colocalized in male rat hypothalamic and limbic neurons that express fos immunoreactivity induced by mating. *Neuroendocrinology* 67, 18-28.
- Gréco, B., Blasberg, M.E., Kosinski, E.C., Blaustein, J.D., 2003. Response of ER α -ir and ER β -ir cells in the forebrain of female rats to mating stimuli. *Horm. Behav.* 43, 444-453.
- Harrington, W.R., Sheng, S., Barnett, D.H., Petz, L.N., Katzenellenbogen, J.A., Katzenellenbogen, B.S., 2003. Activities of estrogen receptor alpha- and beta-selective ligands at diverse estrogen responsive gene sites mediating transactivation or transrepression. *Mol. Cell. Endocrinol.*, 206, 13-22.
- Hillisch, A., Peters, O., Kosemund, D., Muller, G., Walter, A., Schneider, B., Reddersen, G., Elger, W., Fritzemeier, K., 2004. Dissecting physiological roles of estrogen receptor α

and β with potent selective ligands from structure-based design. *Mol. Cell. Endocrinol.* 18, 1599-1609.

Huddleston, G.G., Michael, R.P., Zumpe, D., Clancy, A.N., 2003. Estradiol in the male rat amygdala facilitates mounting but not ejaculation. *Physiol. Behav.* 79, 239-246.

Huddleston, G.G., Paisley, J.C., Clancy, A.N., 2006. Effects of estrogen in the male rat medial amygdala: infusion of an aromatase inhibitor lowers mating and bovine serum albumin conjugated estradiol implants do not promote mating. *Neuroendocrinology* 83, 106-116.

Jensen, E.V., Suzuki, T., Kawashima, T., Stumpf, W.E., Jungblut, P.W., DeSombre, E.R., 1968. A two-step mechanism for the interaction of estradiol with rat uterus. *Proc. Natl. Acad. Sci. USA* 59, 632–638.

Khan M.M., Hadman M., Wakade C., D Sevilla L.M., Dhandapani K.M., Mahesh V.B., Vadlamudi R.K., Brann D.W., 2005. Cloning, expression, and localization of MNAR/PELP1 in rodent brain: colocalization in estrogen receptor alpha but not in gonadotrophin-releasing hormone-positive neurons. *Endocrinology* 146:5215-5227.

Katzenellenbogen, B.S., Katzenellenbogen, J.A., 2000. Estrogen receptor transcription and transactivation: estrogen receptor alpha and estrogen receptor beta: regulation by selective estrogen receptor modulators and importance in breast cancer. *Breast Cancer Research* 2: 335–344.

- King, W.J., Greene, G.L., 1984. Monoclonal antibodies localize oestrogen receptor in the nuclei of target cells. *Nature* 307, 745-747.
- Kirk, R.E., 1968. *Experimental Design: Procedures for the Behavioral Sciences*. Wadsworth Publishing Company, Belmont, California.
- Kuiper, G.G., Enmark, F., Peltö-Huikko, M., Nilsson, S., Gustafson, J.A., 1996. Cloning of a novel receptor expressed in the rat prostate and ovary. *Proc. Natl. Acad. Sci. USA* 93, 5925-5930.
- Kumar, V.M., Vetrivelan, R., Mallick, H.N., 2006. Alpha-1 adrenergic receptors in the medial preoptic area are involved in the induction of sleep. *Neurochem. Res.* 31, 1095-1102.
- Larsson K., Heimer, L., 1964. Mating behavior of male rats after lesions in the preoptic area. *Nature* 202, 413-414.
- Larsson K., Södersten P., Beyer C., 1973. Induction of male sexual behavior by oestradiol benzoate in combination with dihydrotestosterone. *J. Endocrinol.* 57, 563-564.
- Lipton, A., Harvery, H.A., Demers, L.M., Hanagan, J.R., Mulagha, M.T., Kochak, G. M., Fitzsimmons, S., Sanders, S.I., Santen, R.J., 1990. A phase 1 trial of CGS 16949A: a new aromatase inhibitor. *Cancer* 65, 1279-1285.

- Lugg, J.A., Rajfer, J., González-Cadavid, N.F., 1995. Dihydrotestosterone is the active androgen in the maintenance of nitric oxide-mediated penile erection in the rat. *Endocrinology* 136, 1495–1501.
- Macrides, F., Bartke, A., Dalterio, S., 1975. Strange females increase plasma testosterone levels in male mice. *Science* 189, 1104-1106.
- Martini, L., 1982. The 5-alpha reduction of testosterone in the neuroendocrine structures: biochemical and physiological implications. *Endocr. Rev.* 3, 1-25.
- Massa, R., Stupnicka, E., Kniewald, Z., Martini, L., 1982. The transformation of testosterone by the brain and anterior pituitary. *J. Ster. Biochem.* 3, 385-399.
- Mazzucco, C., Walker, H., Pawluski, J., Lieblich, S., Galea, L., 2008. ER α , but not ER β , mediates the expression of sexual behavior in the female rat. *Beh. Brain Res.* 191, 111-117.
- McGinnis, M.Y., Mirth, M.C., 1986. Inhibition of cell nuclear androgen receptors and copulation in male rats by an antiandrogen, Sch 16423. *Neuroendocrinology* 43, 63-68.

McGinnis, M.Y., Dreifuss, R.M., 1989. Evidence for a role of testosterone androgen receptor interactions in mediating masculine sexual behavior in male rats. *Endocrinology* 124, 618-626.

McGinnis, M.Y., Williams, G.W., Lumia, A.R., 1996. Inhibition of male sexual behavior by androgen receptor blockade in the medial preoptic area or the hypothalamus, but not the amygdala or septum. *Physiol. Behav.* 9, 203-213.

Michael, B.J., Vreeburg, T.M., 1973. Copulation in castrated male rats following combined treatment with estradiol and dihydrotestosterone. *Science* 183, 283-285.

Minami, T., Oomura, Y., Nabekura, J., Fukuda, A., 1990. 17β -Estradiol depolarization of hypothalamic neurons is mediated by cyclic AMP. *Brain Res.* 519, 301-307.

Naftolin, F., Ryan, K.J., Davies, I.J., Reddy, V., Flores, F., Petro, Z., Kuhn, M., White, R.J., Takaoka, Y., Wolin L., 1975. The formation of estrogens by central neuroendocrine tissues. *Recent Prog. Horm. Res.* 31, 295-319.

Nomura, M., Korach, K.S., Pfaff, D.W., Ogawa, S., 2003. Estrogen receptor beta (ERbeta) protein levels in neurons depend on estrogen receptor alpha (ERalpha) gene expression and on its ligand in a brain region-specific manner. *Mol. Brain Res.* 110, 7-14.

- Ogaga-Mgbonyebi, E.V., Russell, N.V., Clancy, A.N., 2009. Actions of selective estrogenic drugs in the medial amygdala on male rat mating behavior. Presented at the Annual Meeting of the society for Behavioral Neuroendocrinology, Michigan State University, June 27, 2009.
- Ogawa, S., Lubahn, D.B., Korach, K.S., Pfaff, D.W., 1997. Behavioral effects of estrogen receptor gene disruption in male mice. *Proc. Natl. Acad. Sci. USA* 94, 1476-1481.
- Ogawa, S., Washburn, T.F., Taylor, J., Lubahn, D.B., Korach, K.S., Pfaff, D.W., 1998. Modifications of testosterone-dependent behaviors by estrogen receptor-alpha gene disruption in male mice. *Endocrinology* 139, 5058-5069.
- Ogawa, S., Chester, A.E., Hewitt, S.C., Walker, V.R., Gustafsson, J.Å., Smithies, O., Korach, K.S., Pfaff, D.W., 2000. Abolition of male sexual behaviors in mice lacking estrogen receptors α and β ($\alpha\beta$ ERKO). *Proc. Natl. Acad. Sci. USA* 97, 14737-14741.
- Paisley, J.C., Huddleston, G.G., Denman, H N., Carruth, L L., Grober, M.S., Petrusis, A., Clancy A N., 2005. Inhibition of estrogen receptor synthesis in the medial preoptic area, but not the medial amygdala, reduces male rat mating behavior. *Horm. Behav.* 48, 94.
- Paisley, J.C. 2007. Differential effects of estrogen receptor alpha suppression by antisense oligodeoxynucleotides in the medial preoptic area and the medial amygdala on male rat

mating behavior [Masters Thesis]. Atlanta, GA; Georgia State University.
<http://etd.gsu.edu/theses/available/etd-11302007-154334/>.

Pappas, T.C., Bahiru, G., Watson, C., 1995. Membrane estrogen receptor identified by multiple antibody labeling and impeded-ligand binding. *J. FASEB.* 9, 404–410.

Parte, P., Juneja, H.S., 1992. Temporal changes in the serum levels of gonadotrophins and Testosterone in male rats bearing subcutaneous implants of 5 alpha-dihydrotestosterone. *Int. J. Androl.*, 15, 355–364.

Pietras, R., Szego, C.M., 1977. Specific binding sites for oestrogen at the outer surfaces of isolated endometrial cells. *Nature* 265, 69–72.

Rissman, E.F, Wersinger, S.R., Taylor, J.A., Lubahn, D., 1997. Estrogen receptor functions as revealed by knockout studies: neuroendocrine and behavioral aspects. *Horm. Behav.* 31, 232-243.

Scordalakes, E. M., Imwalle, D. B., Rissman, E. F., 2002. Oestrogen's masculine side: mediation of mating in male mice. *Reproduction* 124, 331-338.

Shughrue, P.J., Merchenthaler, I., 2001. Distribution of estrogen receptor β immunoreactivity in the rat central nervous system. *J. Comp. Neurol.* 436, 64-81.

- Siegel, S., 1956. *Nonparametric Statistics for the Behavioral Sciences*. McGraw Hill Book Company, New York.
- Simerly, R.B. and Swanson, L.W., 1986. The organization of neural inputs to the medial preoptic nucleus of the rat. *J. Comp. Neural.* 246, 312-342.
- Stauffer, S.R., Coletta, C.J., Tedesco, R., Nishiguchi, G., Carlson, K., Sun, J., Katzenellenbogen, B.S., and Katzenellenbogen, J.A., 2000. Pyrazole Ligand: structure-affinity/activity relationship and estrogen receptor- α -selective agonists. *J. Med. Chem.* 43, 4934-4947.
- Sun, J., Ying, R. Huang, Harrington, W.R., Shubin, S., Katzenellenbogen, J.A., Katzenellenbogen, B.S., 2002. Antagonists selective for estrogen receptor. *Endocrinology* 143 , 941-947.
- Swanson, L.W., 1998. *Brain maps: Structure of the Rat Brain*, ed 2, revised. Elsevier, Amsterdam.
- Thomas, P., Pang, Y., Filardo, E.J., Dong, J., 2005. Identity of an estrogen membrane receptor coupled to a G protein in human breast cancer cells. *Endocrinology* 146, 624-632.
- Toufexis, D.J., Myers, K.M., Bowser, M.E., Davis, M., 2007. Estrogen disrupts the inhibition of fear in female rats, possibly through the antagonistic effects of estrogen receptor ER α and ER β . *J. Neurosci.* 7, 9729-9735.

- Vadlamudi, R.K., Brann, D.W., 2005. Cloning, expression, and localization of MNAR/PELP1 in rodent brain: colocalization in estrogen receptor-but not in gonadotropin-releasing hormone-positive neurons. *Endocrinology* 146, 5215–5227.
- Vagell, M.E., McGinnis, M.Y., 1997. The role of aromatization in the restoration of male rat reproductive behavior. *J. Neuroendocrinol.* 9, 415-421.
- Verjans, H.L., van der Molen, H.J., Eik-Nes, K.B., 1975. Relation between circulating levels of testosterone, LH and FSH in intact and castrated, adult, male rats after testosterone administration. *Acta Endocrinol.*, 79, 380-386.
- Walf, A.A., Frye, C.A., 2005. ERbeta-selective estrogen receptor modulators produce antianxiety behavior when administered systemically to ovariectomized rats. *Neuropsychopharmacology* 30, 1598–609.
- Walf, A.A., Ciriza, I., Garcia-Segura, L.M., Frye, C.A., 2007. Antisense oligodeoxynucleotides for estrogen receptor-beta and alpha attenuate estradiol's modulation of affective and sexual behavior, respectively. *Neuropsychopharmacology* epub ahead of print, 2007 Apr 18, PMID:17443129.

Wright, K.D., Cavailles, V., Fuqua, S.A., Jordan, V.C., Katzenellenbogen J.A., Korach, K.S., Maggi, A., Muramatsu, M., Parker, M.G., Gustafsson, J.A., 2006. International Union of Pharmacology. LXIV. Estrogen Receptors. *Pharmacol. Rev.* 58, 773-781.

Zhou, H.B., Carlson, K.E., Stossi, F., Katzenellenbogen, B.S., Katzenellenbogen, J.A., 2009. Analogs of methyl-piperidinopyrazole (MPP): antiestrogens with estrogen receptor alpha selective activity. *Bioorg. Med. Chem. Lett.* 19:108-10.

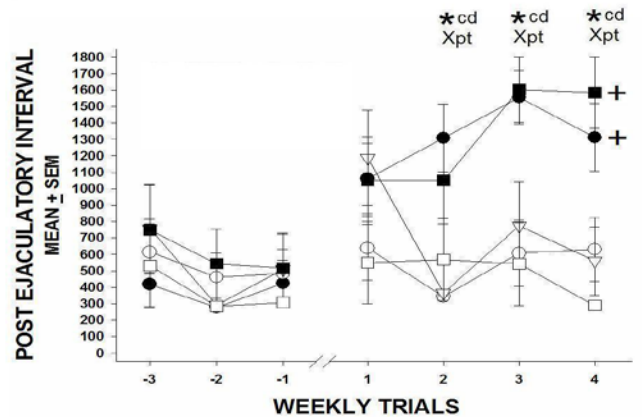
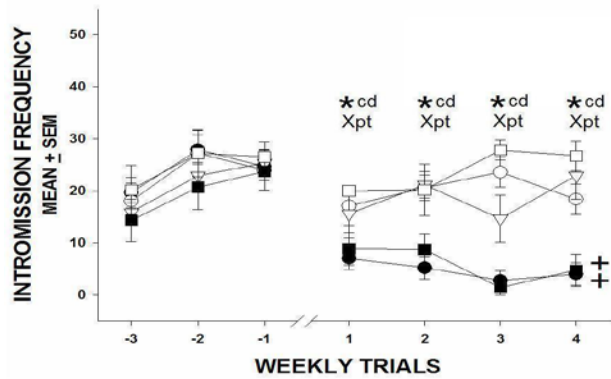
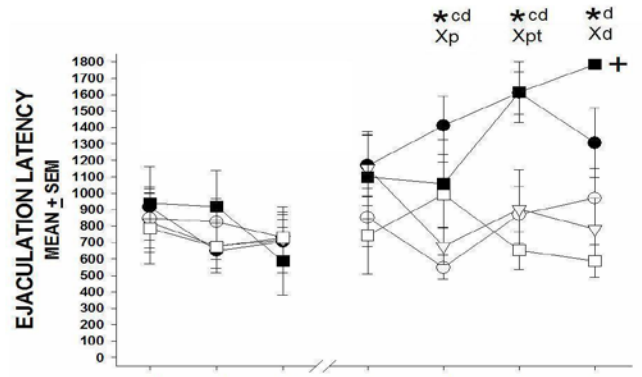
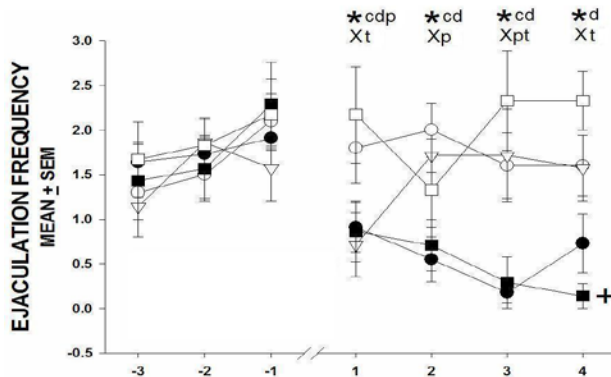
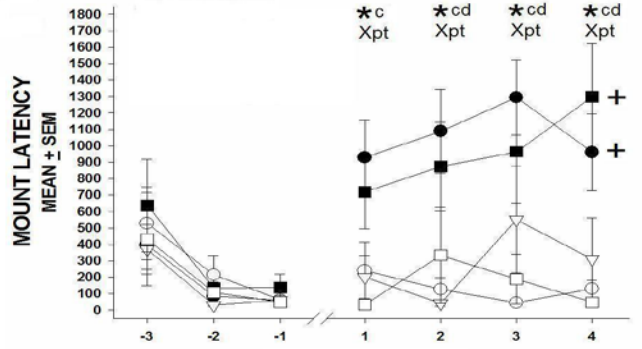
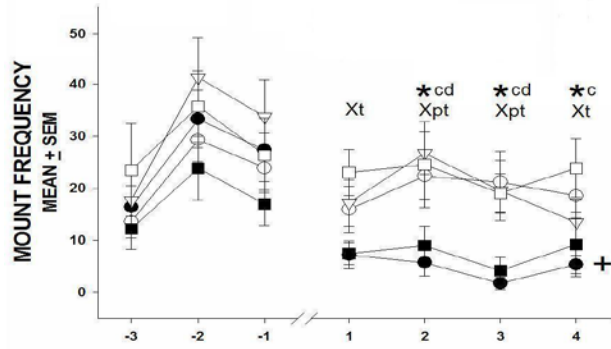


FIGURE 1: MALE SEXUAL BEHAVIOR. Male rats receiving PPT implants to the MPO (p) maintained high levels of mating behavior and did not differ significantly from rats receiving MPO E₂ implants (e). Animals receiving cholesterol MPO implants (c), however, exhibited significantly lower levels of sexual behavior than either of these two groups. DPN did not promote mating in castrated, DHT-maintained males (d) but gonadally intact (DPN toxicity controls) rats (+) mated normally showing that DPN is not toxic.

* Between group difference from estrogen, $p < 0.05$.

X Between group difference from cholesterol, $p < 0.05$.

+ Within group difference from last preoperative sexual behavior test, $p < 0.05$.

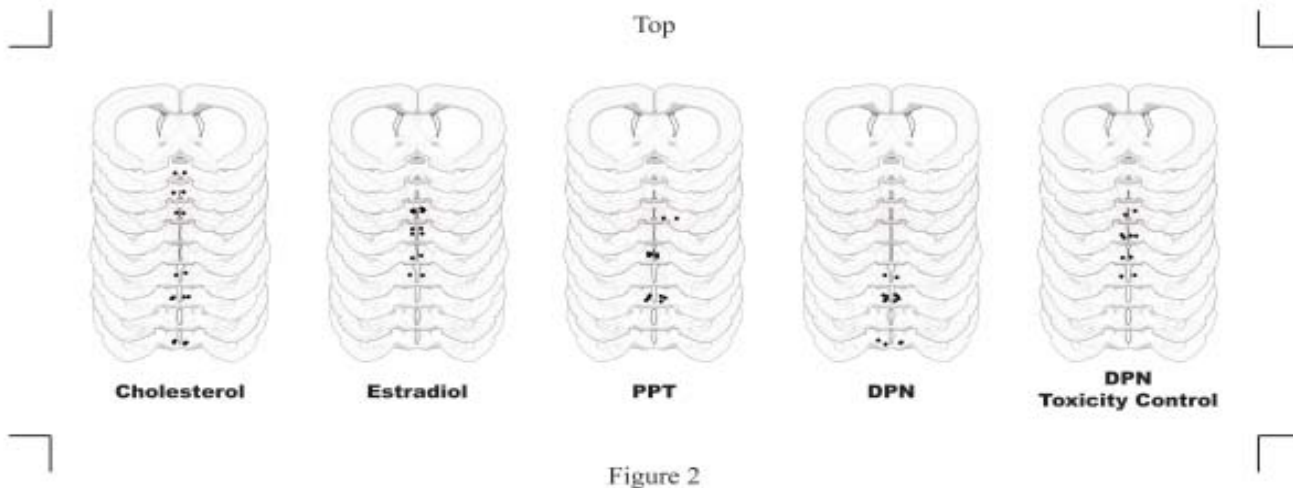


FIGURE 2: EXPERIMENT 1

CANNULA PLACEMENTS TO THE MPO. Cannula tips were located in the MPO or anterior hypothalamus for all males.

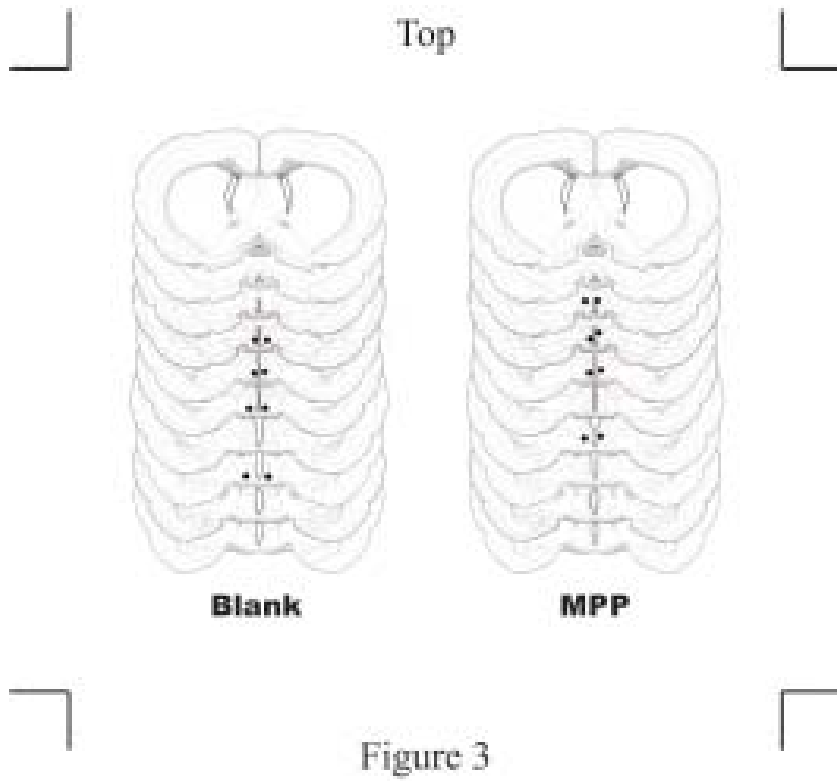


FIGURE 3: EXPERIMENT 2, CANNULA PLACEMENTS TO THE MPO. Cannula tips were located in the MPO or anterior hypothalamus for all males.